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# *Candida albicans* virulence and drug-resistance requires the O-acyltransferase Gup1p

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## Abstract

**Background:** *GUP1* gene was primarily identified in *Saccharomyces cerevisiae* being connected with glycerol uptake defects in association with osmotic stress response. Soon after, Gup1p was implicated in a complex and extensive series of phenotypes involving major cellular processes. These include membrane and wall maintenance, lipid composition, bud-site selection, cytoskeleton orientation, vacuole morphology, secretory/endocytic pathway, GPI anchors remodelling, and lipid-ordered domains assembly, which is compatible with their inclusion in the Membrane Bound O-acyl transferases (MBOAT) family. In mammals, it has been described as a negative regulator of the Sonic hedgehog pathway involved in morphogenesis, differentiation, proliferation, among other processes.

**Results:** We show that *Candida albicans* Gup1p strongly interferes with the capacity of cells to develop hyphae, to adhere, to invade, and to form a biofilm, all of which are significant virulence factors. Furthermore, the mutant colonies exhibited an aberrant morphology/differentiation pattern. Identically to *S. cerevisiae*, *Cagup1Δ* null mutant was more resistant to antifungals like fluconazole, ketoconazole, and clotrimazole, and displayed an abnormal even sterol distribution at the plasma membrane.

**Conclusions:** This work is the first study in the opportunistic yeast *Candida albicans*, showing a role for the *GUP1* gene in virulence as well as in the mechanisms underlying antifungal resistance. Moreover, its impact is even more significant since these results, taken together with all the knowledge about *GUP1* gene (from *S. cerevisiae* and mammals) give consistence to the possibility that Gup1p may be part of a yeast morphogenic pathway parallel to the mammalian Hedgehog.

## Background

*Candida albicans* is a commensal of human microflora, residing at the oral cavity, the gastrointestinal tract, the vaginal and the urinary environments, that acts as an opportunistic pathogen [reviewed by 1]. *C. albicans* commonly causes infections such as denture stomatitis, thrush, and urinary tract-infections, but can also provoke more severe systemic infections. These are frequently life-threatening, in particular in immunocompromised individuals, whose numbers are constantly increasing due to organ transplant, chemotherapy, or, more importantly, to the prevalence of AIDS and Hepatitis C [reviewed by 1]].

Given the limited number of suitable and effective antifungal drugs, together with increasing drug

resistance of the pathogens, it is important that research community addresses, and ultimately discloses, the following yet unsolved questions: a) how the transformation from commensal to pathogen takes place, b) how it can be prevented, c) which are the mechanisms underlying antifungal drugs resistance. All of these culminate in the need to search for new and better agents that target fundamental biological processes and/or pathogenic determinants.

*C. albicans*, as most pathogens, has developed an effective battery of virulence factors and specific strategies to assist the ability to colonize host tissues, cause disease, and overcome host defences [reviewed by [2]]. An outstanding attribute of *C. albicans* biology is its capacity to grow in a diversity of morphological forms, ranging from unicellular budding yeast (blastospores), pseudohyphae, to true hyphae with parallel-sided walls [3-5]. The yeast-hyphae transition contributes to tissue invasion and to the escape from phagocyte cells after

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host internalization [6], and is therefore considered an important virulence factor [4,5,8-11]. Additionally, several other factors have been described in association with virulence, including the production of proteins that mediate adherence, the colonization and invasion of host tissues, the maintenance of cell wall integrity, phenotypic switching, and the avoidance of the host immune response [12-18]. Many of these virulence factors are glycosylphosphatidylinositol (GPI) - anchored proteins, which comprise 88% of all covalently linked cell wall proteins in *C. albicans* [14], many of which associated with the lipid-ordered domains. In spite of all these knowledge, we are still far from fully understanding the precise mechanism(s) driven by *Candida* switch from commensal to pathogen status.

*Saccharomyces cerevisiae* *GUP1* (*ScGUP1*) is a Membrane Bound *O*-acyltransferase (MBOAT) recently proposed to act in the metabolism of lipids, with critical consequences on the lipid-ordered domains stability and assembly [19]. These domains are formed by tight associations of ergosterol and sphingolipids, and aggregate specific proteins, GPI-anchored and non-GPI [19-21]. In accordance, *ScGUP1* has been implicated in the proper GPI-anchors remodelling [22]. Among various classes of lipids in *C. albicans*, membrane ergosterol is an important constituent, which is also the target of common antifungals like polyenes and azoles [23-25]. Therefore, the action of antifungals is affected by changes in the membrane lipid composition, as well as its order (fluidity) and asymmetry in general, and by ergosterol content/distribution in particular [19,23,24,26-28]. Our group has shown [19], that the *Scgup1Δ* mutant displays a moderate sensitivity to sphingolipids biosynthesis inhibitors (SBIs), but a higher resistance to ergosterol biosynthesis inhibitors (EBIs), including azoles. Additionally, the same work shows that the *Scgup1Δ* mutant presents an abnormal sterol distribution in the plasma membrane, as well as internal membranes. In fact, *GUP1* in *S. cerevisiae* has revealed to have a vast pleiotropic nature [19,22,29-32]. In mammals it was described as a negative regulator of the N-terminal palmitoylation of Sonic hedgehog pathway [33], which controls morphogenesis, differentiation and patterning during embryogenesis, including proliferation and cell fate.

In order to explore the involvement of *CaGUP1* in drug susceptibility, we tested the growth of *Cagup1Δ* null mutant in the presence of these compounds. Although, in *C. albicans*, as in *S. cerevisiae*, it is not possible to identify the precise Gup1p acyltransferase dependent reaction/s, we show that the deletion of *GUP1* in *C. albicans* changes ergosterol plasma membrane constitution/distribution, presenting an increased resistance to azoles. More importantly, *CaGup1p*

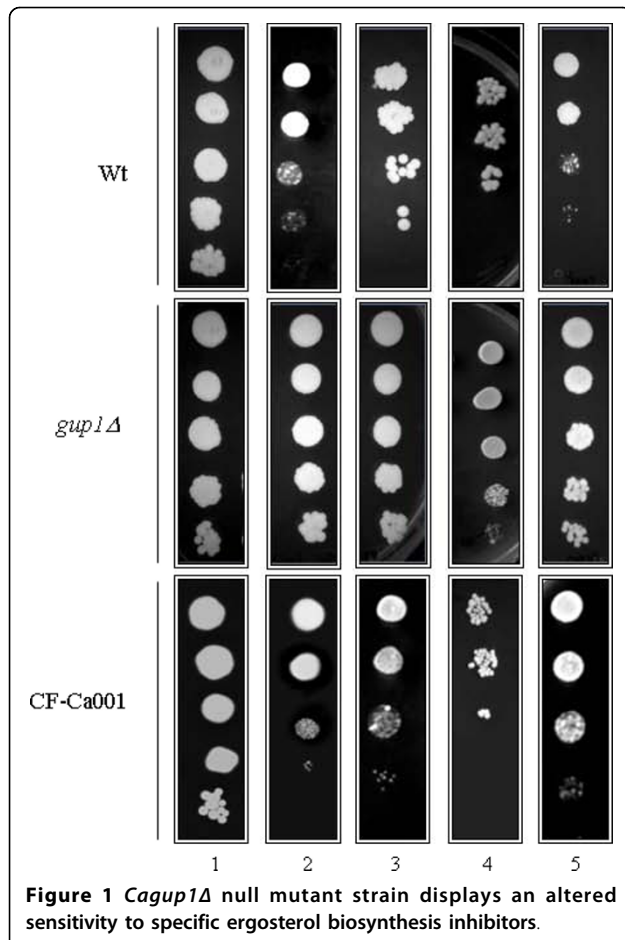
strongly interferes with the capacity of cells to develop hyphae, to adhere, to invade, and to form biofilms, all of which are significant virulence factors. To our knowledge, this work is the first study with *GUP1* gene in *Candida albicans*, and it clearly shows a role for *CaGUP1* gene in virulence.

## Results

### *CaGUP1* deletion provokes resistance to antifungals

The *S. cerevisiae* *O*-acyltransferase Gup1p acts on lipids metabolism affecting the plasma membrane sphingolipids-sterol ordered domains assembly/integrity, and influencing the susceptibility to antifungal drugs [19]. An association between altered lipid-ordered domains and antifungal resistance has been described before [23,24,34,35]. Therefore, we examined the growth behaviour of several clones of *Cagup1Δ* null mutant (3-5) in the presence of some common antifungals and compare them with wt. We used four ergosterol biosynthesis inhibitors (EBIs), hampering different steps of ergosterol biosynthesis [26,27] and two polyenes. The *Cagup1Δ* null mutant strain was equally sensitive to 25 µg/ml amphotericin B and 2.5 µg/ml nystatin as the wt (see Additional file 1). Conversely, *Cagup1Δ* null mutant strain displayed a notorious resistance to all the EBIs used, the azoles with antifungal action, clotrimazole, fluconazole and ketoconazole, and the morpholine fenpropimorph (Figure 1). The resistance of *Cagup1Δ* null mutant strain to clotrimazole and ketoconazole only became obvious at concentrations of 68.8 and 106.3 µg/ml respectively (Figure 1). Moreover, in the presence of 172 µg/ml clotrimazole and of 265.7 µg/ml ketoconazole the growth of both strains was impaired (not shown). The effect of fluconazole, on the other hand, was stronger. The resistance of *Cagup1Δ* null mutant strain to this drug could be detected using 30.6 µg/ml (Figure 1). With regards to fenpropimorph, we verified that, in the presence of 120 and 240 µg/ml of this drug, none of the strains were able to grow (not shown). When the dosage was reduced to 60 µg/ml, the *Cagup1Δ* null mutant strain was more resistant than the parental strain (Figure 1). A copy of the *GUP1* gene, comprising 1.5 Kb of the promoter region and 380 base pairs of the terminator region, was introduced into the *Cagup1* null mutant strain at the *RPS1* locus using the Clp20 plasmid [36]. Correctly, it is possible to see in the same figure that the *GUP1* complemented strain CF-Ca001, displayed a comparable behaviour to wt. Moreover, the introduction of the empty Clp20 plasmid into *Cagup1Δ* null mutant, or into wt, did not cause any amendment on these strains phenotypes (not shown).

Furthermore, we checked if the strains had different growth rates, which could have some impact on these results. Indeed, in liquid medium (which is the only way



**Figure 1** *Cagup1Δ* null mutant strain displays an altered sensitivity to specific ergosterol biosynthesis inhibitors.

Isogenic wt, *Cagup1Δ* null mutant and CF-Ca001 strain were grown to mid-exponential phase in YPD medium. Ten-fold serial dilutions were spotted onto (1) YPD plates (control) and plates supplemented with (2) clotrimazole 68.8 μg/ml, (3) ketoconazole 106.3 μg/ml, (4) fluconazole 30.6 μg/ml and (5) fenpropimorph 60 μg/ml. All plates were incubated at 30°C and photographed after 3-5 days. The *gup1Δ* panel photos are representative of the results obtained with the several clones (3-5) of *Cagup1Δ* null mutant strain tested.

we can compare growth velocities) the doubling time during experimental phase of the wt, mutant and complemented strains is respectively  $1.27 \pm 0.04$  h;  $1.43 \pm 0.06$  h and  $1.25 \pm 0.05$  h. We also determined the mutant doubling time in the presence of fluconazole, which was lower than its value in the absence of the drug. The same happens with the wt. The doubling time during experimental phase of the wt, mutant and complemented strains in the presence of fluconazole are respectively  $1.07 \pm 0.07$  h;  $1.28 \pm 0.09$  h and  $1.11 \pm 0.09$  h.

Alternatively, we used the Methyl-Blue diffusion assay. Consistently, wt strain showed halos with wider diameter than *Cagup1Δ* null mutant strain, indicative that the latter strain is more resistant (see Additional file 2). However, this test provided extra information regarding

the nature of inhibition. The halos displayed by the parental strain were dead-halos, in opposition to growth inhibition halos observed with *Cagup1Δ* null mutant strain (see Additional file 2).

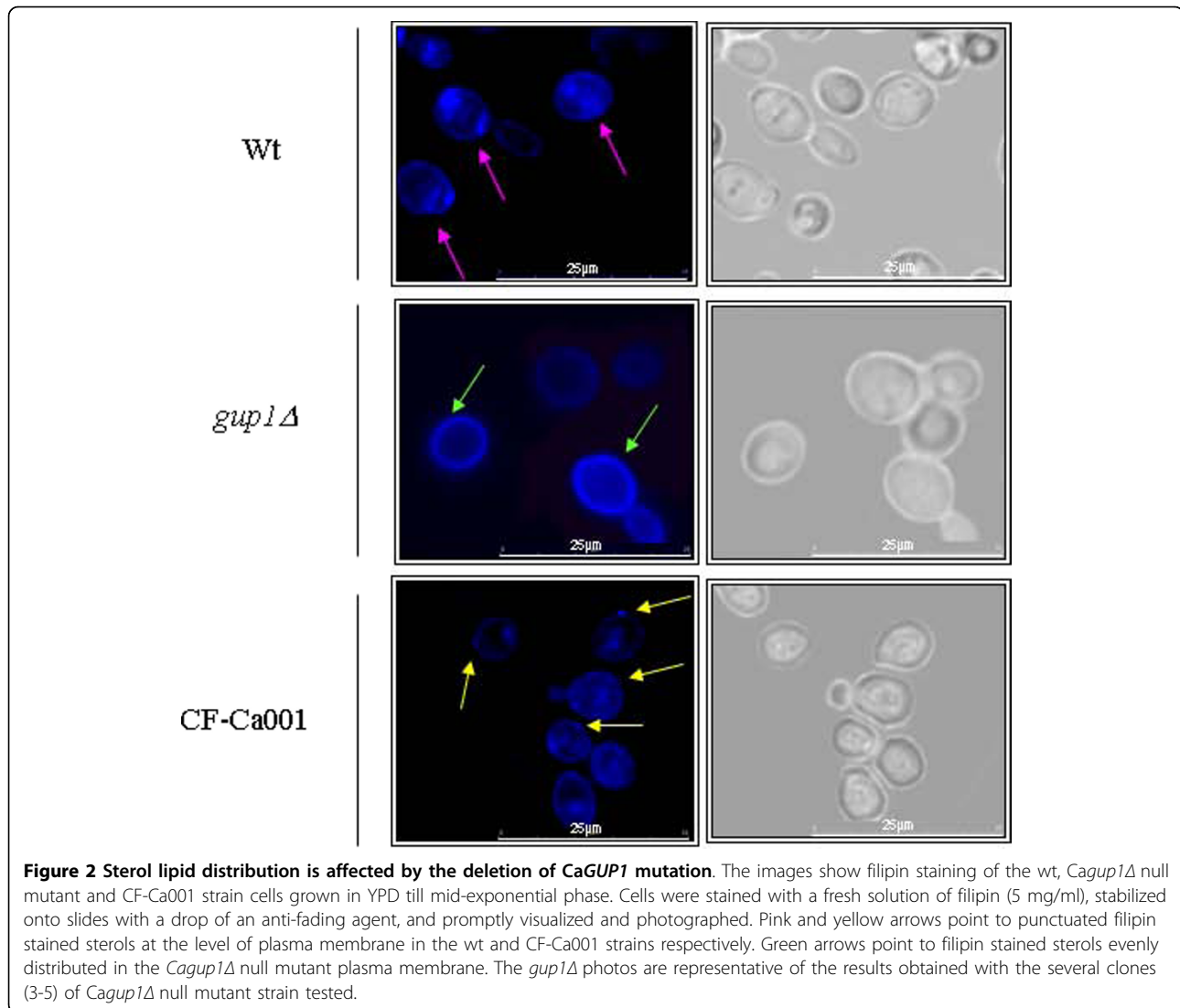
#### ***CaGUP1* deletion affects ergosterol distribution**

The lower susceptibility of the *Cagup1Δ* null mutant strain to antifungals prompted us to analyze ergosterol distribution/occurrence in the plasma membrane. The distribution of free cholesterol in mammalian cells can be visualized by fluorescence microscopy using filipin, a fluorescent antifungal compound that interacts with free 3'-β-hydroxy sterols [37,38]. It has been reported, that the use of filipin needs extra cares. It quickly photobleaches, and given its toxicity, it can deform cell membranes upon a prolonged exposure [19,35,39,40]. These problems were overcome using the optimized method, developed by our group before [19].

The pattern of filipin ergosterol staining on the *Cagup1Δ* null mutant strain differed from the one observed on wt (Figure 2). Overall, fluorescence was mostly present at the cell surface, and *Cagup1Δ* null mutant strain cells were more intensively stained than wt (Figure 2). As expected [19,39-42], the wt plasma membrane was not stained homogeneously, but rather in distinct patches (Figure 2 - pink arrows). In contrast, filipin-stained sterols distributed homogeneously to the *Cagup1Δ* null mutant strain plasma membrane (Figure 2 - green arrows). The complemented strain, CF-Ca001 displayed a pattern of filipin ergosterol staining similar to wt (Figure 2 - yellow arrows). Conversely, the introduction of the empty Clp20 plasmid into the *Cagup1Δ* null mutant, or into wt, did not cause any amendment to these strains phenotypes (not shown). These findings indicate that the maintenance and distribution of normal ergosterol levels in the plasma membrane are altered by *CaGUP1* deletion.

#### **Hypal morphogenesis and colony morphology/ differentiation requires *CaGUP1***

In *C. albicans*, the transition between the yeast form and the filamentous forms has long been a very active area of research, mainly due to its involvement in virulence [reviewed by [4,5,7,10,11]]. Furthermore, *C. albicans*, as well as related species, are able to spontaneously and reversibly make the switch between two or more general phenotypes, reflected by distinct colony morphologies [43]. In order to investigate if *CaGUP1* was implicated in *C. albicans* morphogenesis, young cultures of wt and *Cagup1Δ* null mutant strains were cultivated on agar plates under several conditions. Colonies from both strains formed in non-hypha-inducing conditions (YPD at 30°C) are similar in shape, without peripheral hyphae and no hyphal cells within

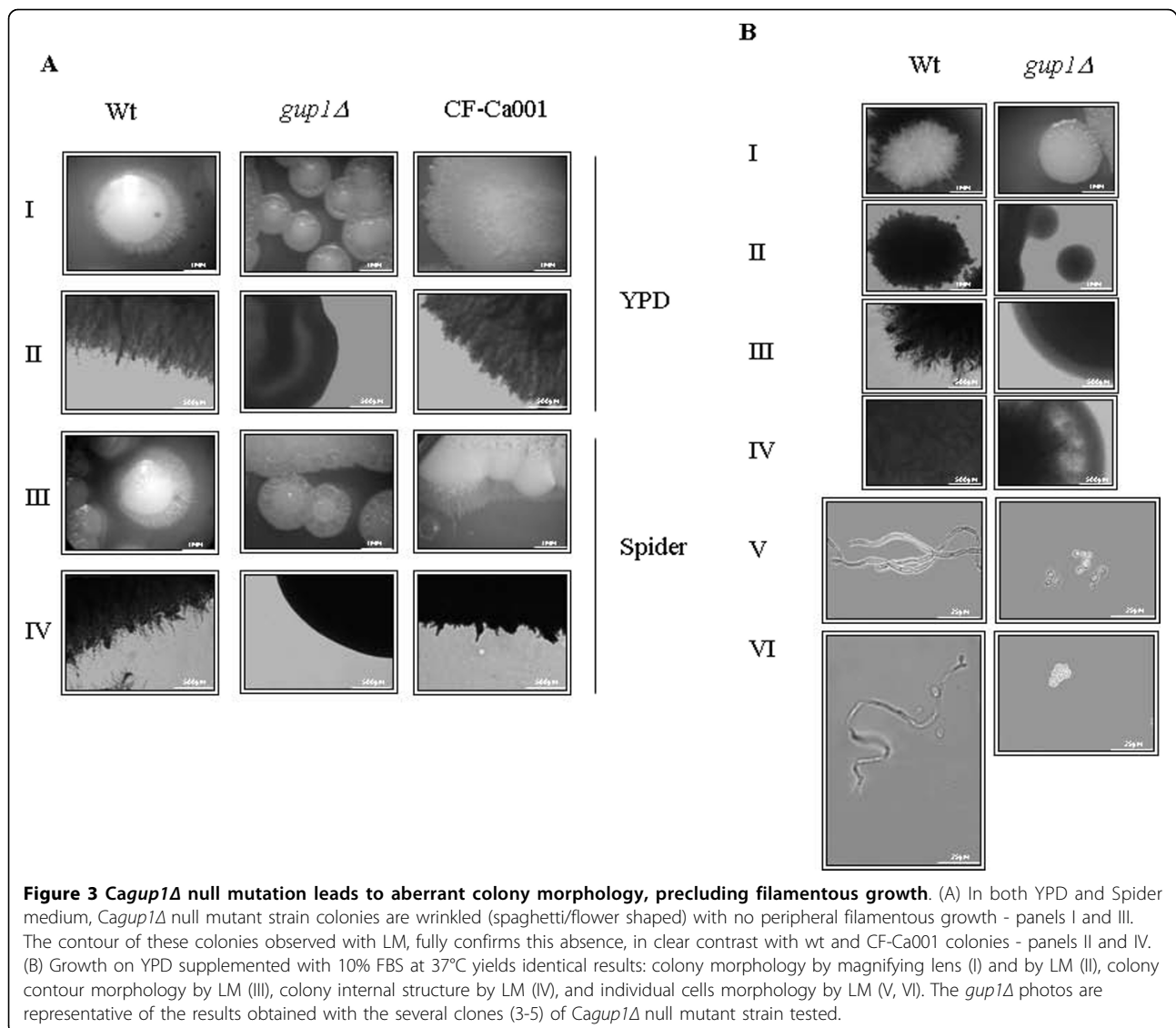


the colony (see Additional file 3). Investigation under hypha-induced conditions presented significant differences between the two strains (Figure 3). In opposition to wt, the colonies of *Cagup1Δ* null mutant strain did not show filaments, either peripheral or inside the colony, suggesting that the mutant lost the ability to form hyphae under the tested conditions. Furthermore, these colonies show a remarkable distinct/aberrant morphology *i.e.* flower, spaghetti, irregular wrinkled shape when compared to wt. In the same figure it is possible to see that, the *GUP1* complemented strain CF-Ca001 displayed a comparable behaviour to wt. The introduction of the empty *Clp20* plasmid into *Cagup1Δ* null mutant or into wt did not cause any amendment on these strains morphology (not shown). Most interesting, when visualized under the microscope, cells within the colonies of the mutant strain were all yeast-type (Figure 3B

- panel V and VI), and not a mixture of hyphae and blastospores as described in the literature [4,44]. The same pattern was observed irrespectively of the medium used.

Time-course of hyphae formation induced by FBS (fetal bovine serum) in liquid medium was also checked. Wt displayed filamentous growth soon after induction (15 min) (Figure 4A) whereas with the *Cagup1Δ* null mutant strain this switch was not observed before 1.5 h. During the remaining time of the experiment, filamentous cells from the *Cagup1Δ* null mutant strain were barely detectable when compared to wt.

Moreover, these filamentous cells were pseudohyphae and not true hyphae as found in wt filamentous cells (Figure 4A, lower panels - time 4 h). Chitin assembly by CFW (Calcofluor white) staining displayed, in the filamentous cells of *Cagup1Δ* null mutant strain,

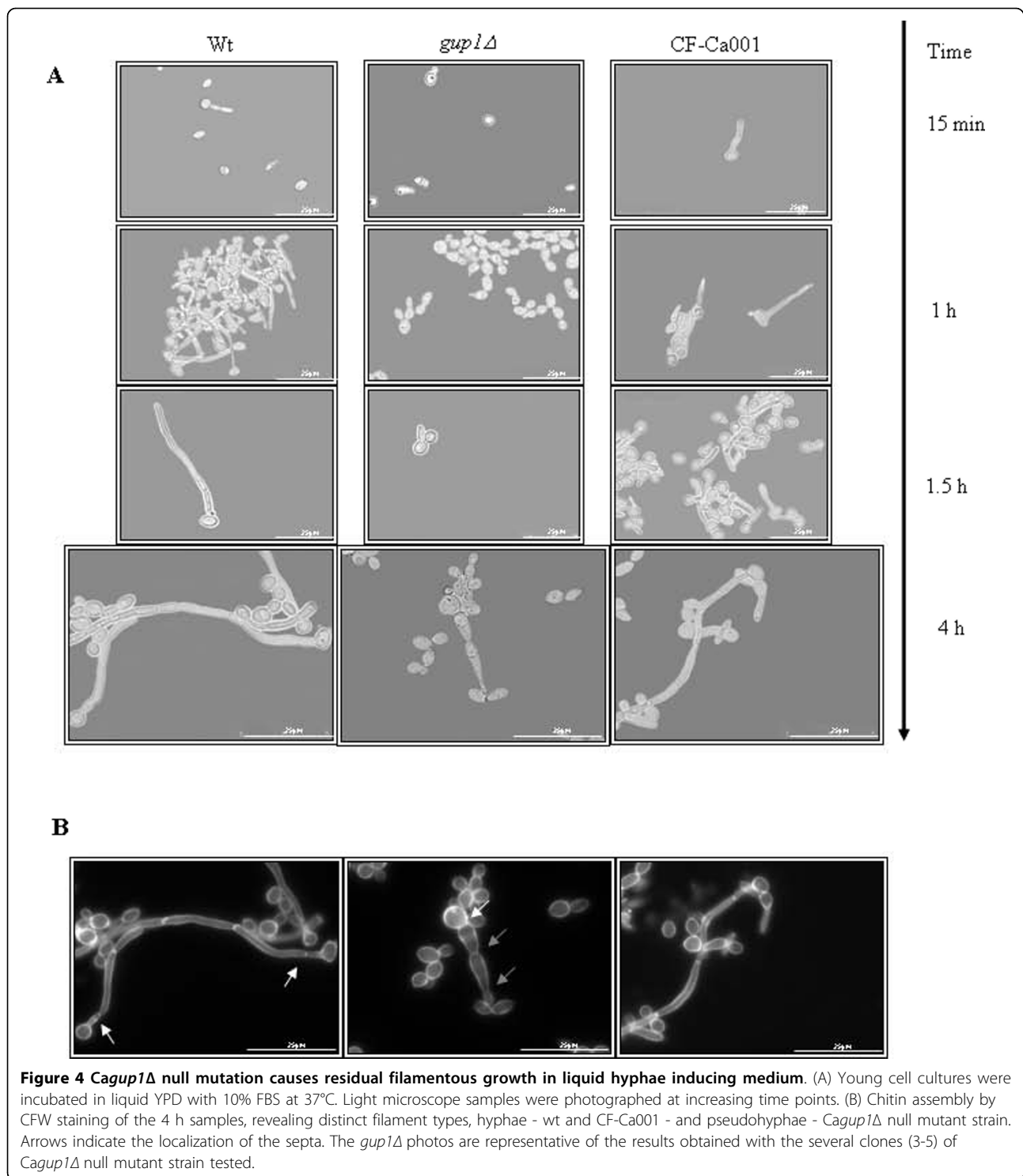


constrictions at the septae junction (Figure 4B - grey arrows) and at the mother-bud neck, where the first septum is located (Figure 4B - white arrows). In opposition, in the wt filamentous cells, which presented true hyphae, the first septum is distant from the mother neck and the other septa do not present constrictions [reviewed by [4] and by [5]]. Additionally, in contrast to wt, in *Cagup1Δ* null mutant strain the elongated compartments were thicker, without parallel sides and were highly branched [reviewed by [4] and [5]]. As before, the *GUP1* complemented strain CF-Ca001, exhibited the same performance as wt (Figure 4), and the control strains with the empty plasmid, act similarly to *Cagup1Δ* null mutant and wt, correspondingly (not shown). These data support the involvement of *CaGUP1* in the morphogenic programme required to induce hyphae

formation, irrespective of the chosen growth regimen (solid or liquid media).

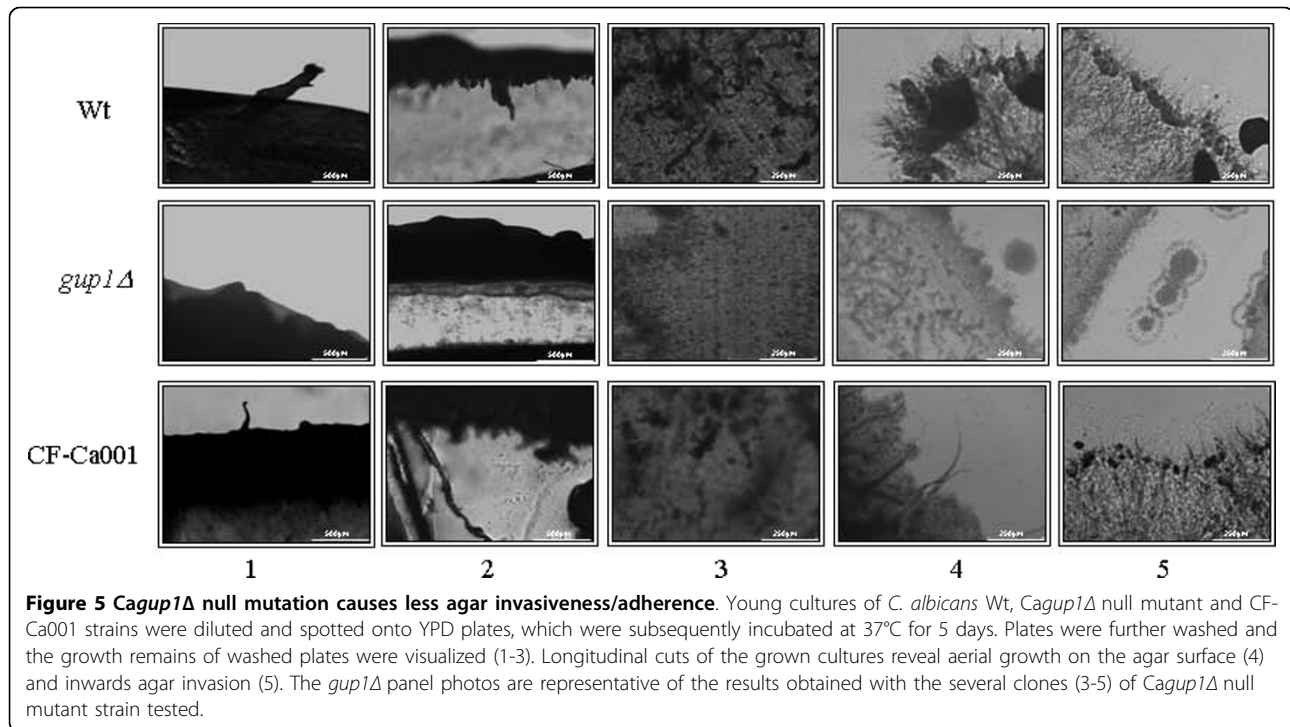
#### Ability of adhesion to polystyrene and invasion of agar is altered on *Cagup1Δ* null mutant

Adhesion of *Cagup1Δ* null mutant strain cells was tested in two different assays: on agar plates with a plate washing assay [45,46], in both YPD and Spider medium, and on polystyrene through the quantification of total biomass by crystal violet (CV) staining [47-49]. The colonies of *Cagup1Δ* null mutant strain were found to be washed away much easier from the agar plates than wt or CF-Ca001 colonies (Figure 5- panels 1-3), indicating that the mutant strain cells have a reduced potential to adhere to the agar. Additionally, microscopic observation of agar surface, as well as



longitudinal cuts revealing the aerial (Figure 5 - panel 4) and inner (Figure 5 - panel 5) agar/growth limits, shows that the wt and CF-Ca001 hyphae extend to aerial environment, but also penetrate/invade the agar (Figure 5 - panel 4-5). Furthermore, these cells which robustly invaded the agar produced hyphae. On the

other hand, the cells of *Cagup1Δ* null mutant strain were not able to penetrate the agar and failed to form hyphae or pseudohyphae. The introduction of the empty *Clp20* plasmid into *Cagup1Δ* null mutant or into wt did not cause any amendment on these strains phenotypes (not shown). These results suggest that the



**Figure 5** *Cagup1Δ* null mutation causes less agar invasiveness/adherence. Young cultures of *C. albicans* Wt, *Cagup1Δ* null mutant and CF-Ca001 strains were diluted and spotted onto YPD plates, which were subsequently incubated at 37°C for 5 days. Plates were further washed and the growth remains of washed plates were visualized (1-3). Longitudinal cuts of the grown cultures reveal aerial growth on the agar surface (4) and inwards agar invasion (5). The *gup1Δ* panel photos are representative of the results obtained with the several clones (3-5) of *Cagup1Δ* null mutant strain tested.

deletion of *CaGUP1* abolishes the ability to invade the agar.

Consonantly, the cells of *Cagup1Δ* null mutant strain also exhibit lower adherence ability to polystyrene (Table 1), comparing to wt and CF-Ca001 cells. This is evidenced by comparing the absorbance values at 2 h incubation time, reflecting the total adhered biomass, corroborated by SEM observation (Figure 6). Light microscopic observation of these samples revealed an amazing lower number of hyphae/pseudohyphae cells on *Cagup1Δ* null mutant strain (not shown). The control strains, with empty plasmid, behaved as expected (not shown).

We also inspect the hydrophobicity of the *Cagup1Δ* null mutant cells, since this factor can influence adhesion. Yet, no significant difference between the % of hydrophobicity of the mutant and wt was observed (2.29% and 2.45% respectively).

**Table 1** Adhesion and Biofilms Assay

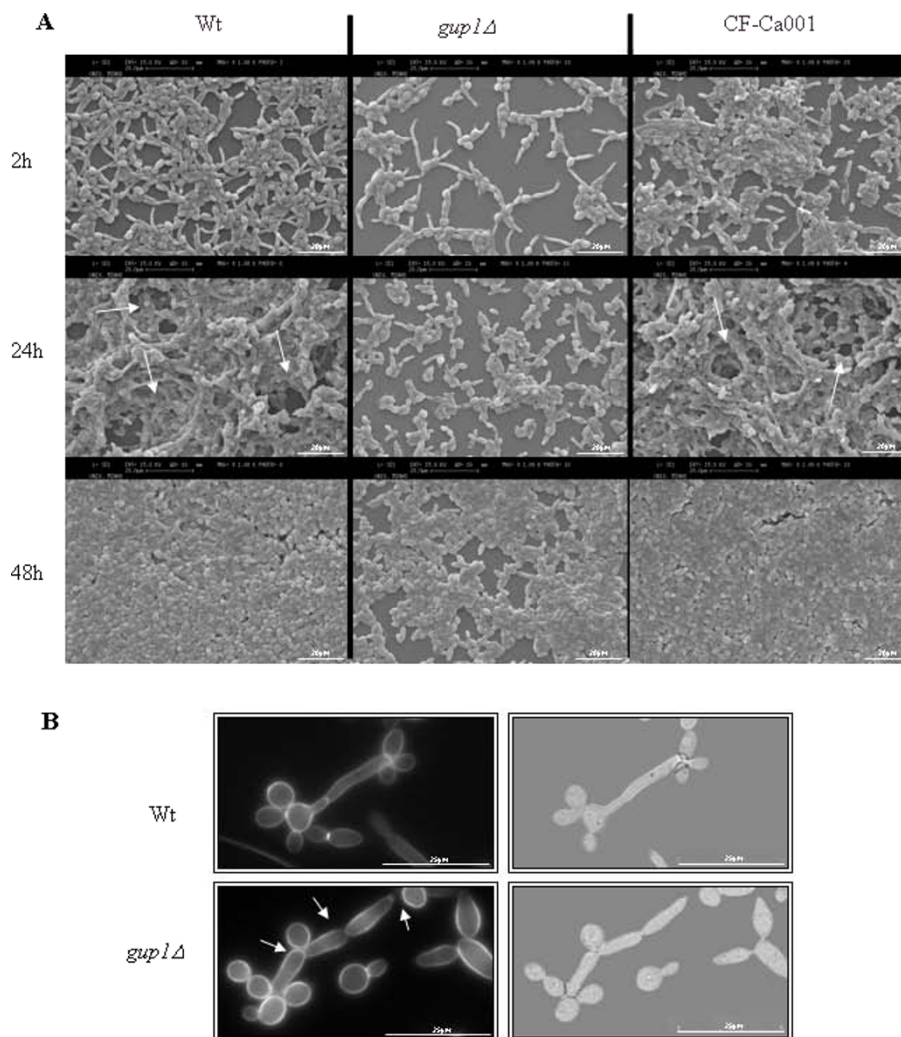
Cell type	Abs values/cm <sup>2</sup> ± SD		
	Time (h)		
	2	24	48
Wt	0.228 ± 0.01	0.324 ± 0.02	0.387 ± 0.06
<i>gup1</i>	0.074 ± 0.01	0.222 ± 0.04	0.293 ± 0.02
CF-Ca001	0.209 ± 0.02	0.298 ± 0.02	0.359 ± 0.04

Standardized absorbance values of Crystal Violet solutions (Abs/cm<sup>2</sup>) obtained in adhesion and biofilms assay of Wt, *Cagup1Δ*, and the control strains ( $\lambda = 570$  nm).

#### Biofilm formation ability is affected in *Cagup1Δ* null mutant

Both filamentation and adhesion of *C. albicans* are involved in the formation of biofilms [50,51], which are commonly found on medical devices, and have attracted attention because of their persistence and resistance to antifungal agents, contributing to both superficial and systemic candidoses [25,50]. We compared the biofilm forming ability of both wt and *Cagup1Δ* null mutant strain cells through the quantification of total biomass by crystal violet (CV) staining [47-49] and Scanning Electron Microscopy (SEM). Importantly, *Cagup1Δ* null mutant strain biofilms had less total biomass compared with wt or with the complemented strain CF-Ca001 (Table 1- absorbance at 24 and 48 h). Wt and the CF-Ca001 strains formed biofilms with biomass  $\approx 1.5$  times higher than the *Cagup1Δ* null mutant strain. The biofilm formation ability of the control strain was as expected. *Cagup1Δ* null mutant strain with the empty Clp20 plasmid, presented the same defect as the mutant and the wt with the empty Clp20 plasmid behaved similarly to wt and the CF-Ca001 (not shown).

SEM observation of the same samples reflected these differences (Figure 6). In opposition to wt or the complemented strain CF-Ca001, *Cagup1Δ* null mutant strain was not able to form typical biofilm structures (Figure 6A). Additionally, *Cagup1Δ* null mutant strain presented much less hyphae/pseudohyphae cells. On the other hand, cell shape inspection by CFW staining (Figure 6B)



**Figure 6** *Cagup1Δ* null mutation causes less and differently structured time-course biofilm formation. (A) SEM micrographs of time course biofilm formation. Arrows indicate the channels observed in a typical biofilm structure - wt and CF-Ca001- not observed in *Cagup1Δ* null mutant strain biofilm. (B) Chitin assembly by CFW staining of individual cells observed by LM. Distinct filament types can be observed. Wt cells display hyphae without septae constrictions, the first septum located within the germ tube, apart from the mother-bud neck (arrow), and less branched, thinner elongated compartments with parallel sides. *Cagup1Δ* null mutant strain cells present pseudohyphae with constrictions located at the septae junctions and at the mother-bud neck, where the first septum is located (arrows), highly branched and thicker elongated compartments without parallel sides. The *gup1Δ* photos are representative of the results obtained with the several clones (3-5) of *Cagup1Δ* null mutant strain tested.

showed that the filamentous cells found in wt biofilm were true hyphae, while the filamentous cells of the *Cagup1Δ* null mutant strain were pseudohyphae (Figure 6B) [4]. As in the induced hyphae experiments (Figure 4), these showed constrictions at the septa and at the mother-bud neck, where the first septum is located, thicker elongated compartments without parallel sides, and highly branched (Figure 6B- white arrows).

## Discussion

In previous works, we showed that *S. cerevisiae* Gup1p, an acyltransferase, is involved in lipids metabolism, with

critical consequences on the plasma membrane lipid-ordered domains stability, on the resistance to antifungals [19], as well as in the cell wall constitution, morphology and assembly [32]. These are important features to be considered when regarding both *C. albicans* switch from commensal to pathogen and its increased resistance to antifungal drugs. Our experiments provide compelling evidence that deletion of both *C. albicans* *GUP1* alleles promotes resistance to antifungals, similarly to what happens in *S. cerevisiae*, but more importantly, CaGup1p interferes in diverse *C. albicans* virulence factors including hyphal development.



Our assumptions are based on the following observations. First, *Cagup1Δ* null mutant strain is resistant to common antifungals. Second, *CaGUP1* deletion provokes an aberrant evenly ergosterol distribution at the level of plasma membrane. Third, the ability to switch from yeast-form to hyphal-growth requires *CaGUP1*. Fourth, a distinct growth orientation elicited by the deletion of *CaGUP1* leads to colonies with remarkable distinct/aberrant morphology *i.e.* flower, spaghetti, irregular wrinkled shape. Fifth, *Cagup1Δ* null mutant strain adherence and invasion abilities are strongly reduced. Sixth, biofilm formation, another important indicator of *C. albicans* virulence, is strongly impaired by the deletion of *CaGUP1*. Finally, the introduction of the *GUP1* gene copy into the *Cagup1Δ* null mutant strain was able to revert all these phenotypes, symptomatic of the *GUP1* gene accountability.

The *C. albicans* laboratory strain BWP17, has recently been subject of great controversy, due not only to the genomic alterations that occurred in its construction, but also due to *URA3* marker [52]. The absence of *URA3* alleles is associated with several phenotypes, some of them regarding *C. albicans* virulence [36,53]. In this work, we were particularly concerned with this, reason why we considered the use of BWP17 as wt control for *GUP1* double deletion as more reliable than the mother strain - SC5314. Both BWP17 and *Cagup1Δ* null mutant present the same genetic background, thus overcoming any possible phenotypic side effects derived from altered chromosomal location of the auxotrophic marker. Furthermore, we introduce the *GUP1* gene copy into the *Cagup1Δ* null mutant strain using Clp20 plasmid [36], since it additionally expresses *URA3* and *HIS1* markers. Integrating vectors are preferable to episomal vectors in *C. albicans*, since they lead to a reduction on the population heterogeneity due to plasmid loss or copy number variance, and this is particularly important for virulence studies. On the other hand, and according to Dennison and co-authors [36], the use of Clp20 plasmid, allows the concomitant regeneration of prototrophy and gene reintegration in null mutants at the *RPS1* locus. Particularly, the integration of *URA3* gene at the *RPS1* locus, circumvent the *URA3* position effects that can complicate the interpretation of *C. albicans* virulence assays [36,52,53]. Finally, two other control strains *Cagup1Δ* null mutant and BWP17 with the empty Clp20 plasmid were constructed, and tested, confirming that the introduction of the empty Clp20 plasmid did not cause any amendment on the mutant or on the wt performance, at any level.

It has been shown that subtle modifications on the membrane lipid composition (phospholipids and ergosterol), on its order (fluidity) and asymmetry could be important determinants of yeast cells susceptibility to

antifungal drugs [23,24,34]. As already referred, *Scgup1Δ* mutant presents a distorted lipidic plasma membrane constitution [54], and a changed stability/assembly of the sphingolipids-sterol ordered domains [19]. Furthermore, in *Scgup1Δ* mutant, ergosterol distribution at the level of plasma membrane is disturbed [19]. As in *S. cerevisiae*, in the *Cagup1Δ* null mutant strain plasma membrane filipin-stained sterols distributed evenly, in contrast with the usual punctuated distribution found in wt plasma membrane. This may be the basis of the observed increased resistance of *Cagup1Δ* null mutant strain to the EBIs, fluconazole, ketoconazole and clotrimazole, belonging to the class of azoles, and the morpholine, fenpropimorph. The azoles are antifungals commonly used to treat yeast infections [23,24,27,28,34]. Although in *C. albicans* the lipid biosynthesis pathways are not well documented, in *S. cerevisiae* these drugs operate on the biosynthesis of ergosterol at the C-14 demethylation stage [27,28], causing a combination of ergosterol depletion and the accumulation of lanosterol, along with other 14-methylated sterols [27,28]. Fenpropimorph, as the other morpholines, inhibits two reactions catalyzed by  $\Delta^{14}$  reductase (an essential enzyme) and  $\Delta^7$ - $\Delta^8$  isomerase [27,28], resulting in the accumulation of 24-methylene ergosterol in the plasma membrane [27,28].

Another group of antifungals, the polyenes, in theory interact specifically with the ergosterol present on the plasma membrane [26,55], creating pores and concomitantly provoking plasma membrane physical and functional disruption, and thus cell death. In spite of the changes observed in ergosterol distribution, *Cagup1Δ* null mutant strain was as sensitive to polyenes as wt. Previous reports, suggest the possibility that polyenes interact also with other membrane lipids besides ergosterol [23,24,34]. In *C. albicans* the metabolism of the other lipids, namely sphingolipids and fatty acids, does not appear to be altered by the deletion of *CaGUP1*, as can be inferred from the susceptibility of the mutant to these lipids biosynthesis specific inhibitors (Ferreira, C., unpublished results).

In a previous work, we found that the absence of *ScGUP1* results in a defective cell wall composition and assembly, with a higher content in  $\beta$ -1,3 glucans and chitin, and lower fraction of mannoproteins [32]. By analogy, and since *C. albicans* and *S. cerevisiae* cell walls are quite alike (with the exception of higher fraction of  $\beta$ -1,6 glucans on the former) [32,56-58], one could consider the possibility of *Cagup1Δ* null mutant cell wall also encompasses higher quantities of  $\beta$ -1,3 glucans. In *C. albicans* it was suggested a correlation between cell wall composition/architecture and resistance to azoles, hypha morphogenesis and virulence [59-61]. Namely, a putative role in azoles resistance on

biofilm cells has been ascribed to  $\beta$ -1,3- glucans [61]. Nett and co-authors described cell wall architectural changes, and increased  $\beta$ -1,3 glucans content associated with fluconazole resistance [61].

Cell wall dynamics in *C. albicans*, underlie regulatory processes during the yeast-to-hyphae transition [59-63]. The ability to switch rapidly between these two forms of growth is a defining characteristic of *C. albicans* cells. Nevertheless, each form of growth provides critical functions required for pathogenicity/virulence [reviewed by [4] and by [5,7]]. Namely, hyphae form is thought to facilitate host tissues invasion and escape from phagocytotic destruction [reviewed by [4] and by [5,7,64]]. We found that, *Cagup1Δ* null mutant strain lost completely the ability to differentiate into hyphae when cultured in solid inducing media conditions, and suffered a considerable delay to grow as filaments when induced with serum in liquid cultures. In this last case, the few remaining *Cagup1Δ* null mutant filamentous cells were smaller, and showed to be pseudohyphae and not true hyphae. When a copy of the *GUP1* gene was introduced into *Cagup1Δ* null mutant, the resulting strain CF-Ca001 regained the ability to differentiate into hyphae, as wt reflecting the role of *GUP1* gene. Interestingly, mammalian *GUP1* gene [33] was able to complement hyphal development defects of *Cagup1Δ* null mutant (Ferreira, C., unpublished results).

The aberrant shape of the *Cagup1Δ* null mutant strain colonies (flower, spaghetti, irregular wrinkled shape) did not present any filamentous cells. This is in accordance with the observed *Cagup1Δ* null mutant defect to grow into hyphae, but appears to be in disagreement with the literature, that attributes a mixture of yeast and hyphae cells to these colonies [reviewed by [4,65,66]]. The complex morphology of these colonies depends, besides other factors, on polarized growth orientation [reviewed by [5,62,63]], which was found to be altered in *Segup1Δ* mutant [30,32]. Additionally, we cannot disregard the possibility that these morphologic cues, may derive from the contribution of the miss-localization/impaired function of specific plasma membrane/wall sensor/proteins.

Invasiveness depends on the existence of hyphae and/or pseudohyphae cells [4]. Accordingly, wt and CF-Ca001 cells were able to invade the agar, whereas *Cagup1Δ* null mutant strain cells lost this ability. This is of extreme relevance in tissue penetration during the early stages of infection. The yeast form might be more suited for dissemination in the bloodstream [4]. Other crucial features with a clear impact on *C. albicans* pathogenicity are the adherence and biofilm formation abilities. The adhesion of *Cagup1Δ* null mutant strain cells either to agar or to polystyrene was greatly reduced when compared to wt and CF-Ca001 strains, which in the former case is in accordance with a lesser agar

invasion, due in part to the lack of filamentous growth. The hydrophobicity of the cells can also influence adhesion, yet *Cagup1Δ* null mutant strain hydrophobicity does not differ from wt. So, their dissimilarities in terms of adherence cannot be explained by this property. However, it is important to highlight that the adhesion phenomenon is not only dependent of cell wall hydrophobicity. Other factors may contribute significantly to it, such as the cell wall charge, cell wall composition (in terms of proteins or other components) [reviewed by [67]] and even the yeast morphology. Moreover, there are many reports acknowledging the inconsistency between the adherence ability and strain hydrophobicity, particularly in *C. albicans* and non-*albicans* isolated strains but also, in other microorganisms as is the case of bacteria [49,68-71].

According to some authors [51,72] three basic stages are necessary for *Candida* biofilm formation *in vitro*: 1) attachment of yeast cells to the surface, 2) growth and proliferation of yeast cells to form a basal layer, followed by 3) extensive filamentation combined with the production of extracellular matrix. We observed a strong defect on the ability of *Cagup1Δ* null mutant strain to form biofilm on an inert substrate (polystyrene wells). The attachment of *Cagup1Δ* null mutant strain cells to this surface, *i.e.* their adherence was nearly one third than the parent strain and no differentiated structure was formed. These observations corroborate defects in the 2 first basic stages above mentioned. Additionally, also the 3<sup>rd</sup>, *i.e.* extensive filamentation was highly compromised.

## Conclusions

In conclusion, we demonstrate that in *Cagup1Δ* null mutant strain the major virulence factors are severely weakened, namely the impaired ability of form true hyphae, to adhere and to invade to different substrates and form biofilms. Equally important, was the revealing role of *CaGUP1* gene in the resistance to antifungals. The present work brings cutting-edge insights into the role of *Gup1p* on the transformation of *C. albicans* into a pathogen. All taken, and considering the fact that *mmGUP1* gene complemented the hyphal morphogenetic defects of *Cagup1Δ* null mutant (Ferreira, C., unpublished results); we anticipate that *Gup1p* may be part of a yeast morphogenic pathway parallel to the mammalian Hedgehog.

## Methods

### Yeast strains, media and growth conditions

*C. albicans* strains used in this work were BWP17 (*ura3Δ::λimm434/ura3Δ ::λimm434his1::hisG/his1::hisGarg4::hisG/arg4::hisG*) [73], several clones (3-5) of homozygous *C. albicans gup1Δ/gup1Δ* (isogenic to

BWP17 but *gup1::URA3-dpl200/gup1::ARG4* [74], and CF-Ca001 (isogenic to *C. albicans gup1Δ/gup1Δ::GUP1*) (this study). All assays were preceded by batch cultures grown on complex medium (YPD: 1% (w/v) yeast extract; 2% (w/v) peptone), supplemented with 2% (w/v) glucose as carbon and energy source, at 26°C to maintain unicellular yeast form. These cultures were continuously inspected as to the absence of hyphae - referred ahead as young cultures. Incubation was done at 160 rpm, orbital shaking with air/liquid ratio 2.5/1. Growth was monitored spectrophotometrically at 600 nm. Solid media were supplemented with 2% (w/v) agar.

Induction of hyphal growth was as follows: Young YPD cultures (above) were inoculated into YPD, YPD + 10% FBS or Spider's medium [1% (w/v) nutrient broth, 1% (w/v) mannitol, 0.2% (w/v) K<sub>2</sub>HPO<sub>4</sub> [75]], supplemented with 1.5% agar, and grown at 37°C for 3-5 days. For time-course induction with FBS in liquid broth, cells from young cultures were washed, resuspended ( $1 \times 10^7$  cell/ml) in YPD supplemented with 10% FBS and incubated at 37°C. Photomicrographs were taken at representative time-points.

#### Strain construction

To reintroduce *GUP1* into *C. albicans gup1Δ/gup1Δ*, the *GUP1* locus was PCR amplified with QIAGEN Long-Range PCR Kit (Qiagen) using the primers ACGCGTCGCTACTTCACATGGTATAAGTGTGCTGATTTGG and GATTAATA TCAAATTTTTCAACCAAGCAAGCATTGAGCTG (MluI and Sali sites underlined), and cloned into the plasmid CIP20 [36] using CloneEZ<sup>®</sup> PCR Cloning Kit (Genscript). CIP20, which is a derivative of CIP10 [76], contains the *URA3* and *HIS1* markers. CIP20-*GUP1* was linearized with StuI, transformed into *C. albicans gup1Δ/gup1Δ* to create the *GUP1*-reintegrant strain CF-Ca001. The integration of CIP20-*GUP1* at the *RPS1* locus was confirmed by PCR with primers TTGTATCACAACCCTCCC and GTGGTTGGAGCTTTGATG. The control strains were generated by transforming the parental strain (BWP17) and the homozygous *C. albicans gup1Δ/gup1Δ* with the empty CIP20 plasmid linearized with StuI.

#### Sensitivity to lipid biosynthesis inhibitors

##### (i) Drop tests

Drop tests were performed from YPD cellular young cultures suspensions, containing approximately  $1 \times 10^6$  cells/ml. Ten-fold serial dilutions were made, and 5 µl of each suspension was applied on the selective media. Results were scored after 3-5 days of incubation at 30°C. Selective conditions were as follow: clotrimazole (68.8 and 172 µg/ml), ketoconazole (106.3 and 265.7 µg/ml), fluconazole (30.6, 91.8 and 153 µg/ml) and fenpropimorph (60, 120 and 240 µg/ml), amphotericin B (25 µg/

ml) and nystatin (2.5 µg/ml). All chemicals were obtained at the highest available grade from Sigma Aldrich.

##### (ii) Methyl-blue diffusion test

Alternatively, we assayed the sensitivity to lipid biosynthesis inhibitors with a methyl-blue-diffusion plate test. Sterile filter disks (BBL) of 6 mm diameter were placed on top of YPD methyl-blue plates seeded with 5 ml of a wt or *Cagup1Δ* mutant strain young cultures. The filter disks were impregnated with 5 to 10 µl of the following drugs: clotrimazole (137.6 µg/ml), ketoconazole (212.6 µg/ml), fluconazole (91.8 µg/ml), fenpropimorph (80 µg/ml), amphotericin B (25 µg/ml) and nystatin (2.5 µg/ml). The plates were incubated at 30°C, and halos of inhibition were scored after 3 days. Again, all chemicals were obtained at the highest available grade (Sigma-Aldrich).

#### Filipin/Sterol fluorescence microscopy

Sterol-lipid distribution was assessed *in vivo* using filipin. This was performed basically as described before [19,40]. For fluorescence microscopy, cells were mounted directly on slides with a 10 µl drop of anti-fading agent Vectashield (Vector Laboratories) to overcome the instability of filipin, and immediately observed by light microscopy (LM).

#### Colony morphology and differentiation

To observe different colony morphology/differentiation, equal volumes of young cultures of each strain were diluted and spotted onto non-inducing (YPD at 30°C) and hyphal-inducing (Spider medium and on 10% FBS at 37°C) conditions, and also in YPD at 37°C. Cultures were allowed to grow for 3-5 days. Colonies on agar surface were observed under magnifying lens (10 times) and photographed. Spider medium colonies were also thoroughly observed by light microscopy. To visualize cellular morphology within the colony, cells of each colony phenotype were resuspended and washed twice in PBS. Approximately 10 µl of the suspensions were then mounted on glass slides and cells were visualized by LM.

#### Chitin assembly analysis

To discriminate between hyphae and pseudohyphae cell wall chitin assembly was assessed with CFW staining. Cultures were diluted to  $1 \times 10^7$  cells/ml and to 1 ml of cells suspension was added 100 µl of CFW (300 µg/ml). Samples were incubated at room temperature for 5 min and 5 µl of each suspension placed on glass slide for microscopic inspection. The dye fluoresces when bound to chitin, primarily, and to glucans, staining cell wall and septa. Representative images were obtained by LM.

### Adherence to agar and invasion capacities

Equal volumes of young cultures of each strain were diluted to  $1 \times 10^7$  cells/ml, and 1 ml of cells suspension was spotted onto YPD medium agar plates. Solid cultures were allowed to grow at 37°C for 5 days. The cells on the surface were removed by washing under running water [45,46] and then visualized by LM. Inspection of agar invasion was performed by visualization of longitudinal cuts displaying the aerial and internal agar/growth boundaries by LM.

### Light microscopy

Microscopy assessments were done in a Leica Microsystems DM-5000B epifluorescence microscope, with appropriate filter settings. Images were acquired through a Leica DCF350FX digital camera and processed with LAS AF Leica Microsystems software.

### Cell wall hydrophobicity

MATH test was utilized to evaluate cell wall hydrophobicity as described by Rosenberg [77]. Yeast cells were harvested in stationary phase and washed twice with PBS pH 7.0. A yeast cell suspension displaying an optical density at 600 nm ( $OD_{600\text{ nm}}$ ) between 0.4–0.5 was prepared in PBS ( $A_0$ ). In an acid washed spectrophotometer glass tubes, 3 ml of the prepared yeast suspension was spread and overlaid by 0.4 ml of a hydrophobic hydrocarbon, hexadecane. After vigorous vortexing, phases were allowed to separate for 10 min at 30°C and  $OD_{600\text{ nm}}$  of the aqueous phase was measured ( $A_1$ ). The percentage of hydrophobicity was calculated as follows: hydrophobicity (%) =  $[1 - (A_1/A_0)] \times 100$ . Assays were performed in triplicate and statistical analysis (T-test,  $p < 0.05$ ) of the results was carried out.

### Adhesion and biofilm formation

Adhesion and biofilm formation ability was assessed through quantification of total biomass by crystal violet (CV) staining as described before [47–49]. For this, standardized cell suspensions (1 ml containing  $1 \times 10^7$  cells/ml in YPD) from young cultures were placed into selected wells on polystyrene plates (Orange Scientific, Braine-l'Alleud, Belgium) and incubated at 37°C in a shaker at 120 rev/min. Adhesion ability was measured after 2 h of incubation and biofilm formation ability was inspected after 24 h and 48 h. Regarding the 48 h sample, an extra step was performed, at half period, *i.e.* at 24 h, 500  $\mu$ l of YPD medium were removed and an equal volume of fresh YPD added. After the defined times of incubation, the medium was aspirated and non-adherent cells removed by washing the wells with sterile ultra-pure water. Following, the adherent cells were fixed with 1 ml of methanol, which was removed after 15 min of contact. The plates were allowed to dry at

room temperature, and 1 ml of CV (1% v/v) was added to each well and incubated for 5 min. The wells were then gently washed with sterile, ultra-pure water, and 1 ml of acetic acid (33% v/v) was added to release and dissolve the stain. The absorbance of the obtained solution was read at 570 nm in triplicate in a microtiter plate reader (Bio-Tek Synergy HT, Izasa, Lisbon, Portugal). The final absorbance was standardized according to the volume of acetic acid and area of the wells ( $\text{abs}/\text{cm}^2$ ). Three to five independent assays were performed for each experiment.

### Scanning electron microscopy

Structure of adhered and/or biofilm cells were examined by Scanning Electron Microscopy (SEM). For this, medium and non-adherent cells were extracted as described for CV staining (above). Samples were then dehydrated with alcohol (using 70% ethanol for 10 min; 95% ethanol for 10 min and 100% of ethanol for 20 min) and air dried for 20 min. The bases of the wells were cut and were kept in a desiccator until analysed. Samples were then covered with gold for visualization in a S-360 scanning electron microscope (Leo, Cambridge, USA).

### Additional material

**Additional file 1: Growth inhibition halos in the presence of polyenes.** Sterile filter disks were impregnated with 25  $\mu$ g/ml amphotericin B (AmpB) and 2.5  $\mu$ g/ml nystatin (Nys) and placed on top of YPD methyl-blue plates seeded with 5 ml of a wt or *Cagup1Δ* null mutant strain mid-exponential phase cultures. Halos of growth inhibition were measured (mm) after 2 or 3 days.

**Additional file 2: Growth inhibition halos in the presence of EBIs.** Sterile filter disks were impregnated with the drugs and placed on top of YPD methyl-blue plates seeded with 5 ml of a wt or *Cagup1Δ* null mutant strain mid-exponential phase cultures. (1) YPD plates (control) and plates with the impregnated disks (2) clotrimazole 137.6  $\mu$ g/ml, (3) ketoconazole 212.6  $\mu$ g/ml, (4) fluconazole 91.8  $\mu$ g/ml and (5) fenpropimorph 80  $\mu$ g/ml. Halos of growth inhibition were measured (mm) after 2 or 3 days.

**Additional file 3: Colony morphology under non-hypha-inducing conditions.** *Cagup1Δ* null mutant and wt present similar colony morphology in non-hypha-inducing conditions. Equal volumes of young cultures of each strain were diluted and spotted onto YPD, and allowed to grow at 30°C for 3–5 days.

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#### Authors' contributions

CF conceived the project, performed EBI's sensibility, filipin, hyphal development, morphology, microscopy (light and SEM), adhesion and invasion abilities experiments and wrote the paper; SS and MH conceived and performed biofilm experiments; SS also participate in SEM experiments; EP and FFO performed complementation experiments; CL provided the founds and executed critical readings of the manuscript at several stages. All authors have read and approved the final manuscript.

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#### References

- Pfaller MA, Diekema DJ: **Epidemiology of invasive candidiasis: a persistent public health problem.** *Clin Microbiol Rev* 2007, **20**:133-163.
- Naglik JR, Challacombe SJ, Hube B: **Candida albicans secreted aspartyl proteinases in virulence and pathogenesis.** *Microbiol Mol Biol Rev* 2003, **67**:400-428.
- Gow NA, Brown AJ, Odds FC: **Fungal morphogenesis and host invasion.** *Curr Opin Microbiol* 2002, **5**:366-371.
- Sudbery P, Gow N, Berman J: **The distinct morphogenic states of Candida albicans.** *Trends Microbiol* 2004, **12**:317-324.
- Whiteway M, Bachewich C: **Morphogenesis in Candida albicans.** *Annu Rev Microbiol* 2007, **61**:529-553.
- Kumamoto C, Vines M: **Contributions of hyphae and hyphae-co-regulated genes to Candida albicans virulence.** *Cell Microbiol* 2005, **7**:1546-1554.
- Brown AJ: **Morphogenetic Signalling Pathways in Candida albicans.** In *Candida and Candidiasis*. Edited by: Calderone RA. ASM Press, Washington DC; 2002:95-106.
- Lo HJ, Kohler JR, DiDomenico BB, Loebenberg D, Cacciapuoti A, Fink GR: **Nonfilamentous C. albicans mutants are avirulent.** *Cell* 1997, **90**:939-949.
- Mitchell AP: **Dimorphism and virulence in Candida albicans.** *Curr Opin Microbiol* 1998, **1**:687-692.
- Saville SP, Lazzell AL, Monteagudo C, Lopez-Ribot JL: **Engineered control of cell morphology in vivo reveals distinct roles for yeast and filamentous forms of Candida albicans during infection.** *Eukaryot Cell* 2003, **2**:1053-1060.
- Saville SP, Lazzell AL, Bryant AP, Fretzen A, Monreal A, Solberg EO, Monteagudo C, Lopez-Ribot JL, Milne GT: **Inhibition of filamentation can be used to treat disseminated Candidiasis.** *Antimicrob Agents Chemother* 2006, **50**:3312-3316.
- Fu Y, Luo G, Spellberg BJ, Edwards JE, Ibrahim AS: **Gene overexpression/suppression analysis of candidate virulence factors of Candida albicans.** *Eukaryot Cell* 2008, **7**:483-492.
- Hube B, Sanglard D, Odds FC, Hess D, Monod M, Schafer W, Brown AJ, Gow NA: **Disruption of each of the secreted aspartyl proteinase genes SAP1, SAP2, and SAP3 of Candida albicans attenuates virulence.** *Infect Immun* 1997, **65**:3529-3538.
- Kapteyn JC, Hoyer LL, Hecht JE, Muller WH, Andel A, Verkleij AJ, Makarow M, Van den Ende H, Klis FM: **The cell wall architecture of Candida albicans wild-type cells and cell wall-defective mutants.** *Mol Microbiol* 2000, **35**:601-611.
- Leidich SD, Ibrahim AS, Fu Y, Koul A, Jessup C, Vitullo J, Fonzi W, Mirbod F, Nakashima S, Nozawa Y, Ghannoum MA: **Cloning and disruption of caPLB1, a phospholipase B gene involved in the pathogenicity of Candida albicans.** *J Biol Chem* 1998, **273**:26078-26086.
- Romani L, Bistoni F, Puccetti P: **Adaptation of Candida albicans to the host environment: the role of morphogenesis in virulence and survival in mammalian hosts.** *Curr Opin Microbiol* 2003, **6**:338-343.
- Sanglard D, Hube B, Monod M, Odds FC, Gow NA: **A triple deletion of the secreted aspartyl proteinase genes SAP4, SAP5, and SAP6 of Candida albicans causes attenuated virulence.** *Infect Immun* 1997, **65**:3539-3546.
- Sheppard DC, Yeaman MR, Welch WH, Phan WH, Fu QT, Ibrahim Y, Filler AS, Zhang SG, Waring M, Edwards AJ, Edwards JE: **Functional and structural diversity in the Als protein family of Candida albicans.** *J Biol Chem* 2004, **279**:30480-30489.
- Ferreira C, Lucas C: **The yeast O-acyltransferase Gup1p interferes in lipid metabolism with direct consequences on the sphingolipid-sterol-ordered domains integrity/assembly.** *Biochim Biophys Acta* 2008, **1778**:2648-2653.
- Edidin M: **The state of lipid-rafts: from model membranes to cells.** *Annu Rev Biophys Biomol Struct* 2003, **32**:257-283.
- Jacobson K, Mouritsen OG, Anderson RG: **Lipid rafts: at a crossroad between cell biology and physics.** *Nat Cell Biol* 2007, **9**:7-14.
- Bosson R, Jaquenoud M, Conzelmann A: **GUP1 of Saccharomyces cerevisiae encodes an O-acyltransferase involved in remodelling of the GPI anchor.** *Mol Biol Cell* 2006, **17**:2636-2645.
- Pasirja R, Krishnamurthy S, Prasad T, Ernst J, Prasad R: **Squalene epoxidase encoded by ERG1 affects morphogenesis and drug susceptibilities of Candida albicans.** *J Antimicrob Chemother* 2005, **55**:905-913.
- Pasirja R, Prasad T, Prasad R: **Membrane raft lipid constituents affect drug susceptibilities of Candida albicans.** *Biochem Soc Trans* 2005, **33**:1219-1223.
- Sanglard D, Ischer F, Parkinson T, Falconer D, Bille J: **Candida albicans mutations in the ergosterol biosynthetic pathway and resistance to several antifungal agents.** *Antimicrob Agents Chemother* 2003, **47**:2404-2412.
- Georgopapadakou NH, Walsh TJ: **Antifungal agents: chemotherapeutic targets and immunologic strategies.** *Antimicrob Agents Chemother* 1996, **40**:279-291.
- Song Z, Nes WD: **Sterol biosynthesis inhibitors: potential for transition state analogs and mechanism-based inactivators targeted at sterol methyltransferase.** *Lipids* 2007, **42**:15-33.
- Warringer J, Blomberg A: **Involvement of yeast YOL151W/GRE2 in ergosterol metabolism.** *Yeast* 2006, **23**:389-398.
- Holst B, Lunde C, Lages F, Oliveira R, Lucas C, Kielland-Brandt MC: **GUP1 and its close homologue GUP2, encoding multimembrane-spanning proteins involved in active glycerol uptake in Saccharomyces cerevisiae.** *Mol Microbiol* 2000, **37**:108-124.
- Ni L, Snyder M: **A genomic study of the bipolar bud site selection pattern in Saccharomyces cerevisiae.** *Mol Biol Cell* 2001, **12**:2147-2170.
- Bonangelino CJ, Chavez EM, Bonifacio JS: **Genomic screen for vacuolar protein sorting genes in Saccharomyces cerevisiae.** *Mol Biol Cell* 2002, **13**:2486-2501.
- Ferreira C, Silva S, von Voorst F, Aguiar C, Kielland-Brandt MC, Lucas C, Brandt A: **Absence of Gup1p in Saccharomyces cerevisiae results in a defective cell wall composition, assembly, stability and morphology.** *FEMS Yeast Res* 2006, **6**:1027-1038.
- Abe Y, Yoshiko K, Niikura T: **Mammalian Gup1, a homologue of Saccharomyces cerevisiae glycerol uptake/transporter 1, acts as a negative regulator for N-terminal palmitoylation of Sonic hedgehog.** *FEBS J* 2008, **275**:318-331.
- Mukhopadhyay K, Prasad T, Saini P, Pucadyil J, Chattopadhyay A, Prasad R: **Membrane sphingolipid-ergosterol interactions are important determinants of multidrug resistance in Candida albicans.** *Antimicrob Agents Chemother* 2004, **48**:1778-1787.
- Cánovas D, Pérez-Martin J: **Sphingolipid biosynthesis is required for polar growth in the dimorphic phytopathogen Ustilago maydis.** *Fungal Genet Biol* 2009, **46**:963-975.
- Dennison PM, Ramsdale M, Manson CL, Brown JP: **Gene disruption in Candida albicans using a synthetic, codon-optimised Cre-loxP system.** *Fungal Genet Biol* 2005, **42**:737-748.
- Norman AW, Demel RA, de Kruijff B, van Deenen LLM: **Studies on the biological properties of polyene antibiotics. Evidence for the direct interaction of filipin with cholesterol.** *J Biol Chem* 1972, **247**:1918-1929.
- Severs NJ: **Cholesterol cytochemistry in cell biology and disease.** *Subcell Biochem* 1997, **28**:477-505.
- Alvarez FJ, Douglas LM, Konopka JB: **Sterol-rich plasma membrane domains in Fungi.** *Eukaryot Cell* 2007, **6**:755-763.
- Grossmann G, Opekarova M, Malinsky J, Weig-Meckl I, Tanner W: **Membrane potential governs lateral segregation of plasma membrane proteins and lipids in yeast.** *EMBO J* 2007, **26**:1-8.
- Beh CT, Rine J: **A role for yeast oxysterol-binding protein homologs in endocytosis and in the maintenance of intracellular sterol-lipid distribution.** *J Cell Sci* 2004, **117**:2983-2996.
- Takeda T, Chang F: **Role of fission yeast myosin I in the organization of sterol-rich membrane domains.** *Curr Biol* 2005, **15**:1331-1336.

43. Zhao R, Lockhart SR, Daniels K, Soll DR: **Roles of TUP1 in switching, phase maintenance, and phase-specific gene expression in *Candida albicans*.** *Eukaryot Cell* 2002, **1**:353-365.
44. Laffey SF, Butler G: **Phenotype switching affects biofilm formation by *Candida parapsilosis*.** *Microbiol* 2005, **151**:1073-1081.
45. Guo B, Styles CA, Feng Q, Fink GR: **A *Saccharomyces* gene family involved in invasive growth, cell-cell adhesion, and mating.** *Proc Natl Acad Sci USA* 2000, **97**:12158-12163.
46. Hube B, Hess D, Baker CA, Schaller M, Schafer W, Dolan JW: **The role and relevance of phospholipase D1 during growth and dimorphism of *Candida albicans*.** *Microbiol* 2001, **147**:879-889.
47. Stepanovic S, Vukovic D, Dakic I, Savic B, Svabic-Vlahovic M: **A modified microtiter-plate test for quantification of staphylococcal biofilm formation.** *J Microbiol Meth* 2000, **2**:175-179.
48. Henriques M, Azeredo J, Oliveira R: ***Candida albicans* and *Candida dubliniensis*: comparison of biofilm formation in terms of biomass and activity.** *Brit J Biomed Sci* 2006, **63**:5-11.
49. Silva S, Henriques M, Martins A, Oliveira R, Williams D, Azeredo J: **Biofilms of non-*Candida albicans* *Candida* species: quantification, structure and matrix composition.** *Med Mycol* 2009, **20**:1-9.
50. Hiller E, Heine S, Brunner H, Rupp S: ***Candida albicans* Sun41p, a putative glycosidase, is involved in morphogenesis, cell wall biogenesis, and biofilm formation.** *Eukaryot Cell* 2007, **6**:2056-2065.
51. Nobile CJ, Mitchell AP: **Genetics and genomics of *Candida albicans* biofilm formation.** *Cell Microbiol* 2006, **8**:1382-1391.
52. Selmecki A, Bergmann S, Berman J: **Comparative genome hybridization reveals widespread aneuploidy in *Candida albicans* laboratory strains.** *Mol Microbiol* 2005, **55**:1553-1565.
53. Brand A, MacCallum DM, Brown AJP, Gow NA, Odds FC: **Ectopic expression of URA3 can influence the virulence phenotypes and proteome of *Candida albicans* but can be overcome by targeted reintegration of URA3 at the RPS10 locus.** *Eukaryot Cell* 2004, **3**:900-909.
54. Oelkers P, Pinkelenberg A, Erdeniz N, Cromley D, Billheimer J, Sturley S: **A lecithin cholesterol acyltransferase-like gene mediates diacylglycerol esterification in yeast.** *J Biol Chem* 2000, **275**:15609-15612.
55. Silva L, Coutinho A, Fedorov A, Prieto M: **Nystatin-induced lipid vesicles permeabilization is strongly dependent on sterol structure.** *Biochim Biophys Acta* 2006, **1758**:452-459.
56. Klis FM, de Groot P, Hellingwerf K: **Molecular organization of the cell wall of *Candida albicans*.** *Med Mycol* 2001, **39**:1-8.
57. Klis FM, Mol P, Hellingwerf K, Brul S: **Dynamics of cell wall structure in *Saccharomyces cerevisiae*.** *FEMS Microbiol Rev* 2002, **26**:239-253.
58. Netea MG, Gow NA, Munro CA, Bates S, Collins C, Ferwerda G, Hobson RP, Bertram G, Hughes HB, Jansen T, Jacobs L, Buurman ET, Gijzen K, Williams DL, Torensma R, McKinnon A, MacCallum DM, Odds FC, van der Meer JW, Brown AJ, Kullberg BJ: **Immune sensing of *Candida albicans* requires cooperative recognition of mannans and glucans by lectin and Toll-like receptors.** *J Clin Invest* 2006, **116**:1642-1650.
59. Angioletta L, Micoci MM, D'Alessio S, Girolamo A, Maras B, Cassone A: **Identification of major glucan-associated cell wall proteins of *C. albicans* and their role in fluconazole resistance.** *Antimicrob Agents Chemother* 2002, **1688**-1694.
60. Herrero AB, Magnelli P, Mansour MK, Levitz SM, Bussey H, Abeijon C: ***KRES* gene null mutant strains of *Candida albicans* are a virulent and have altered cell wall composition and hyphae formation properties.** *Eukaryot Cell* 2004, **3**:1423-1431.
61. Nett J, Lincoln L, Marchillo K, Massey R, Holoyda K, Hoff B, van Handel , Andes D: **Putative role of  $\beta$ -1,3 Glucans in *Candida albicans* biofilm resistance.** *Antimicrob Agents Chemother* 2007, **51**:510-520.
62. Oberholzer U, Marcil A, Leberer E, Thomas DY, Whiteway M: **Myosin I is required for hypha formation in *Candida albicans*.** *Eukaryot Cell* 2002, **1**:213-228.
63. Oberholzer U, Iouk TL, Thomas DY, Whiteway M: **Functional characterization of myosin I tail regions in *Candida albicans*.** *Eukaryot Cell* 2004, **3**:1272-1286.
64. Zheng X, Wang Y, Wang Y: **Hgc1, a novel hypha-specific G1 cyclin-related protein regulates *Candida albicans* hyphal morphogenesis.** *EMBO J* 2004, **1845**-1856.
65. Slutsky B, Buffo J, Soll DR: **High-frequency switching of colony morphology in *Candida albicans*.** *Science* 1985, **230**:666-669.
66. Soll DR: **Phenotypic switching.** In *Candida and Candidiasis*. Edited by: Calderone RA. ASM Press, Washington DC; 2002:123-142.
67. Brown AJ, Odds FC, Gow NA: **Infection-related gene expression in *Candida albicans*.** *Curr Opin Microbiol* 2007, **10**:307-313.
68. Cerca N, Pier GB, Vilanova M, Azeredo J: **Quantitative analysis of adhesion and biofilm formation on hydrophilic and hydrophobic surfaces of clinical isolates of *Staphylococcus epidermidis*.** *Res Microbiol* 2005, **156**:506-514.
69. Henriques M, Oliveira R, Azeredo J: **The involvement of physico-chemical interactions in the adhesion of *Candida albicans* and *Candida dubliniensis* to epithelial cells.** *Mycoses* 2007, **50**:391-396.
70. Silva S, Teixeira P, Oliveira R, Azeredo J: **Adhesion to and viability of *Listeria monocytogenes* on food contact surfaces.** *J Food Protect* 2008, **71**:1379-1385.
71. Sousa C, Henriques M, Teixeira P, Oliveira R: **Influence of surface properties on the adhesion of *Staphylococcus epidermidis* to acrylic and silicone.** *Int J Biomater* 2009.
72. Chandra J, Kuhn DM, Mukherjee PK, Hoyer LL, McCormick T, Ghannoum MA: **Biofilm formation by the fungal pathogen *Candida albicans*: development, architecture and drug resistance.** *J Bacteriol* 2001, **183**:5385-5394.
73. Wilson RB, Davis D, Mitchell AP: **Rapid hypothesis testing in *Candida albicans* through gene disruption with short homology regions.** *J Bacteriol* 1999, **181**:868-874.
74. Kayingo G, Martins A, Andrie R, Andrie R, Neves L, Lucas C, Wong B: **A permease encoded by *STL1* is required for active glycerol uptake by *Candida albicans*.** *Microbiol* 2009, **155**:1547-1557.
75. Liu H, Hohler J, Fink GR: **Suppression of hyphal formation in *Candida albicans* by mutation of *STE12* homolog.** *Science* 1994, **266**:1723-1726.
76. Murad AM, Lee PR, Broadbent ID, Barell CJ, Brown AJ: **Clp10, an efficient and convenient integrating vector for *Candida albicans*.** *Yeast* 2000, **16**:325-327.
77. Rosenberg M: **Bacterial adherence to hydrocarbons: a simple method to measure cell-surface hydrophobicity.** *FEMS Microbiol Lett* 1980, **22**:289-295.

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